

Review



## RECENT UPDATES OF SILK BASED NOVEL DRUG DELIVERY SYSTEMS

Narinder Singh\*, Surya Prakash Gautam, Shalini Dyal, Lovepreet Singh, Harjaskaran.

Department of Pharmaceutics, CT institute of Pharmaceutical Sciences, Shahpur, Jalandhar.

Submitted On: 25.03.2016

Revised On: 28.03.2016

Accepted On: 31.03.2016

### ABSTRACT

Silk is natural protein basically comprised of fibroin and sericin. Major resource of silk is Oak tasar silkworm *Antheraea proylei*, Muga silkworm *Antheraea assamensis*, tasar silkworm *Antheraea proylei*, mulberry silkworm *Bombyx mori* and Eri silkworm *Philosamia ricini*. Special features like mechanical toughness, self-assembly, biocompatible, biodegradability and processing flexibility or low immunogenicity proteins are offering their utility for gene and drug delivery. Silk Fibroin based nanoparticles are prepared by techniques like electrospinning, salting out, supercritical fluid technology, desolvation and mechanical comminution. Each method has a significance important in preparation of Silk Fibroin based nanoparticles for drug delivery. Silk fibroin has broadly used 3D-silk fibroin scaffolds, microspheres, films, nanoparticles, Nano fibers, biomedical sutures, for coatings, sponges, biomaterials for implants and tubes as well as for drug delivery. Patents for silk based drug delivery applications have attracted the researcher involved in drug delivery area.

**KEY-WORDS:** Silk Worms, Silk Proteins, preparation methods,

Corresponding Author: **Narinder Singh**  
Phone Number: +91 9464394755  
E-mail: [Pharmacist.narinder@gmail.com](mailto:Pharmacist.narinder@gmail.com)

Indian Research Journal of Pharmacy and Science; 8(2016) 456-470  
Journal home page: <https://www.irjps.in>

**INTRODUCTION:**

Silk being a natural protein fiber which controlled generally of fibroin and made by certain insect larvae to form cocoons. These silk protein molecules retain bio-degradable, bio-compatible and self assembling properties which are natively improved with gene delivery or offering utility for drug and genetic engineering. Some properties of silk-based material like solubility, biodegradability, and mechanical properties, can be managed by employing the secondary structure. Innovative engineering techniques together with mild all-aqueous progressions consume expanded range of submission to nucleic acid therapeutics and complex protein. Silk attained from silkworm is used in biomedical sutures for decades<sup>1,2</sup>. SF matrices were confirmed to positively deliver protein drugs and reserve their effectiveness. Silk fibroin have been recommended for delivery on the technique via drug delivery inside the usage of hydrogels porous 3D scaffolds and films. The best characterization silks are the cocoon silk cultivated from the silkworm dragline silk from the spider *Bombyx mori* and *Nephila clavipes*.<sup>3,4</sup> Targeted delivery can be attained Hybrid and merged silk-based materials comprising some biopolymers, which has not been widely studied. It should deliver applicable mechanical, or natural possessions for not only drug delivery and gene delivery. It has also intended for regenerative medicine or medical imaging and tissue engineering. Silk are characterized in three types<sup>5</sup>. Silk I, water soluble which have composed of combination of  $\alpha$ -helix and  $\beta$ -turn structures, and random coil, Silk II, categorized by a majority of  $\beta$ -sheet which are stable and water insoluble fibroin; Silk III, considered as  $\alpha$ -helix and usually found at the water or air interface.<sup>6,7</sup> Silk fibroin encompass exceptional potential as a carrier

for sustained or controlled release of cargo. Silk has importance used role in human body in the form of suture material.

**Types of Silk Fibroins****1. Silkworm Silk fibroins**

Silk fibroin is used as biomedical sutures from ancient time. It is also been used in textile production for clothing, due to the fact that silkworms are easier to domesticate.<sup>8</sup> The core progression in the heavy chain contain the alanine-glycine repeats. In the silkworm cocoons, two fibroins are sheathed in a glue-like, sericin coat proteins, to form the composite fibers of the cocoon. The mainly premeditated silkworm silk proteins, contain two major components, heavy chain (~325 kDa) and light (~25 kDa) fibroins. Various methods are now used to extract and regenerate silk fibroin. Several silk based biomaterials, such as silk films, porous scaffolds electrospun Nanofibers, and hydrogels can be processed from silk solutions.<sup>9</sup>

**2. Spider Silk Fibroins**

*Nephila clavipes* is the most universal and commonly deliberate spider silk in terms of structure. Function is dragline silk which is veiled as a mixture of two proteins from focused columnar epithelial cells of the most important ampullate secretor of weaver spinning spiders. The molecular weights of Silk proteins range from 70 to 700 kDa depending on source. Partial cDNA clones programming the two types of dragline silks had been secluded and analyze from two species of orb-web weaving spiders, *N. clavipes* (MaSpI and MaSpII) and *Araneus diadematus* (ADF-3 and ADF4).<sup>10</sup> The silks proteins are characterized for block copolymers, tranquil of large hydrophobic blocks with highly

preserved repetitive in order consisting of small side-chain amino acids, like alanine and glycine, with prevailing short hydrophilic block with extramultipart progression. Which have been prepared via amino acids with charged amino acids and larger side chain. The hydrophobic blocks form beta-sheets and physically cross-linked crystalline domains in silk fibers. The imposing tensile strength of silk fibers is due to the presence of hydrophobic or less ordered hydrophilic regions. Silk fibers are combination with chain direction achieved during revolving.<sup>11</sup>

### TYPES OF SILKWORM

There are five types of silk collected from altered species of silkworms:-

#### 1) Mulberry

The large of the commercial silk are formed cutting-edge in world originates after the diversity and often silk generally famous as mulberry silk. Mulberry silk are obtained from the silkworm, *Bombyx mori*<sup>12,13</sup> These silkworms are entirely cultivated and reared inside. In India, the foremost mulberry silk cultivated states are, West Bengal, Karnataka, Tamil Nadu, Andhra Pradesh, Andhra Pradesh, and Jammu & Kashmir.<sup>14</sup>

#### 2) Tasar

Tasar (Tussah) is the copperish colour, silk mostly used for fittings and interiors. It is fewer shiny than mulberry silk. Tasar silk is generated by the silkworm, *Antheraea mylitta* which mostly increase on food plants Asan and Arjun. In India, tasar silk is largely formed in the states of Chhattisgarh, Orissa, and Jharkhand, besides Maharashtra, Andhra Pradesh and West Bengal. Tasar culture is the key continue for lots of a ethnic community in India.<sup>15</sup>

#### 3) Oak Tasar

It is a greater diversity of tasar generated by the silkworm, *Antheraea proylei*. In India which are provided for natural food trees of oak, initiate in profusion in the sub-Himalayan tie of India covering the states of Himachal Pradesh, Jammu & Kashmir, Assam, Uttar Pradesh, Meghalaya, China and Manipur. China is the most imperative inventor of oak tasar in the world. Tasar derives from altered silkworm which is known as *Antheraea pernyi*.<sup>15</sup>

#### 4) Eri

It's also well-known as Errandi or Endi, Eri has a multivoltine silk spin from open-ended cocoons, nothing similar further variations of silk. Eri silk is the product of the cultivated silkworm, *Philosamia ricini* that feeds mostly on castor leaves. Eri society is a domestic activity proficient mostly for delicacy for the tribal and protein rich pupae. The silk is worn native for production of *chaddars* (wraps) for have used by these tribals. In India, society is expert mostly in the Assam and north-eastern states.<sup>10,16</sup>

#### 5) Muga

Muga is golden yellow colour silk which are privilege of India and the pleasure state of Assam. It is collected from semi-cultivated multivoltine silk *Antheraea assamensis*. These silkworms forage on the fragrant leaves of Soaluplant and are raised on trees similar to that of tasar. Muga silk is mostly cultivated in the state of Assam and basic part of the tradition. The muga silk as high value creation is used in products like Machala's and sarees.<sup>12,17</sup>

### RECOMBINANT SILK PROTEINS

In the previous few years ago, many methods had been used in thoughtful silk inheritance, structures and biophysics cloning. Silk proteins personalized via

genetic engineering can also be planned to present original features at the side of native properties.<sup>18</sup>

### 1. Silkworm Variant

Silkworm silk is obtained from *B. morisilkworm* & elastin obstruct copolymers or silk elastin similar to proteins constructed by recombinant DNA techniques. Silk had been utilized as gene and drug release systems forming hydrogels to liberate adenovirus containing writer genes increase the cell-adhesive facility of silk-fibroin partial collagen and fibronectin sequences. Silk fibroin from natural silkworm *Anaphe* has a large amount simpler amino acid composition in evaluation to *B. morisilkworm*

silk fibroin. *Anaphe* silk fibroin may be a suitable for the design and formulation for the recombinant proteins.<sup>19,20</sup>

### 2. Spider Variants

A spider silk sequence was customized to have methionines adjacent to the polyalanine (beta sheet forming domain) sequence and derived from the silaffin protein of the diatom *Cylindrothecafusiformis*. Silk based amphiphilic have been developed to improve the transfection efficiency through integrin-mediated endocytosis. The designs can be extensive to further control targeting, stability and size or related some needs for gene delivery.<sup>21</sup>

**Table No 1.** Comparison of silk proteins from the cocoons of mulberry silkworm *B.mori*

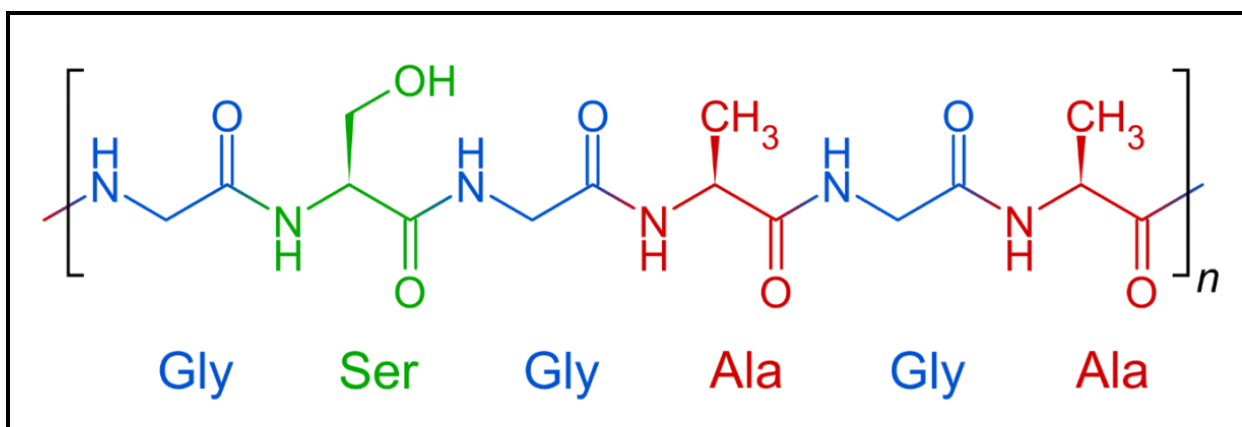
Protein	Fibroin	Sericin
<b>Composition</b>	Amino acids in a heavy chain and a light chain in a 1:1 ratio linked by disulfide bonds. <sup>22</sup>	Polypeptide polymer consisting of 18 amino acids <sup>23</sup>
<b>Type of protein</b>	Structural protein	Glue-like protein
<b>Proportion of cocoon</b>	70%	20-30%
<b>Structural Properties</b>	Hydrophobic; $\beta$ -sheets and $\alpha$ -helices	Hydrophilic; remains in a partially unfolded state, with high proportion of random coil structure <sup>23</sup>
<b>Molecular weight</b>	26-370 kDa	24-250 kDa
<b>Major properties</b>	Biocompatible, Biodegradable, Crystallinity, Mechanical Properties, Opportunity for Chemical Functionalization <sup>24</sup>	Antibacterial, UV-absorbing, high Moisture Absorbancy, Antioxidant, Antitumor, Wound-healing <sup>25</sup>
<b>Novel applications</b>	Drug delivery, Tissue engineering, Implant coating, imaging and diagnostics <sup>24</sup>	Food, Cosmetics, Drug delivery, Medical and Pharmaceutical Industries
<b>Advantages for use in drug delivery</b>	Aqueous processing, controllable biodegradation	Capable of carrying both hydrophilic and hydrophobic drugs

**PROPERTIES OF SILK FIBRES.<sup>26,27</sup>**

**Table No 2. Physical & Chemical Properties of Silk Fibers.**

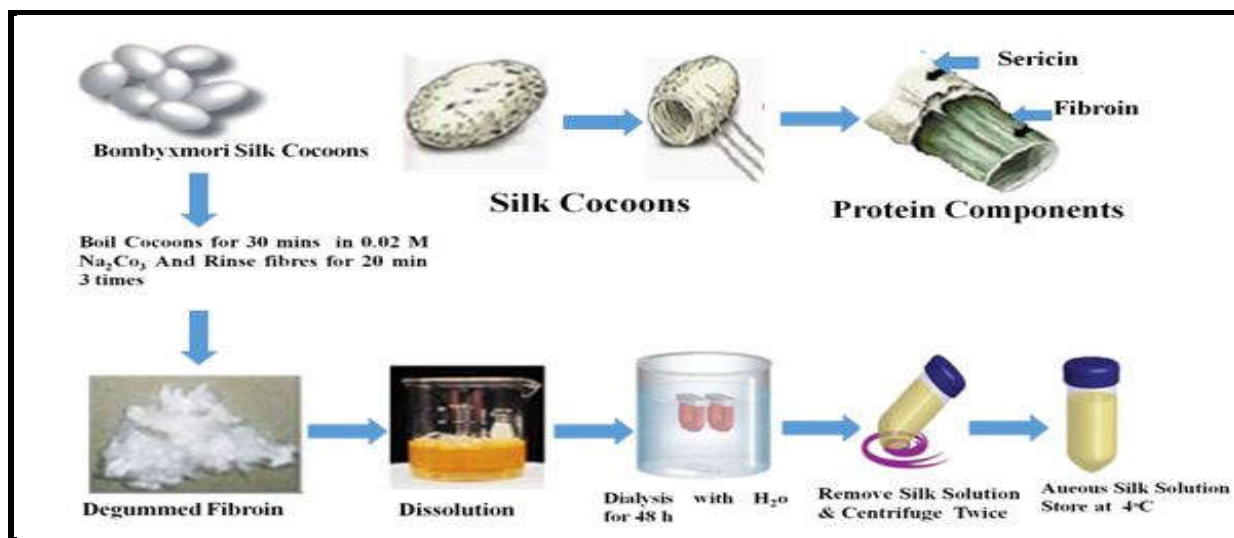
Physical properties	Chemical Properties :
Silkworm includes a triangular cross section shape with curved corners, 5-10 µm wide. The Fibroin made up of generally beta-sheets, due to presence a 59 amino acid repeat sequence with several variations.	Silk is opposed to mainy mineral acids, apart from for sulfuric acid, in which dissolve it.
Silk have numerous synthetic fibers, unlike soft texture that is not slippery, and smooth.	Silkworm consists of mainly two main proteins fibroin and sericin.
Silk has susceptible to static cling and poor conductor of electricity.	The high percentages (50%) of glycine, which are few amino acid, allow stiff stuffing. Fibers are opposed to breaking and tough.
It have also used in favor of the denier and measurement of linear density in fibers	Fibroin consist of the heavy amino acids chains Gly-Ser-Gly-Ala-Gly-Ala and developed beta sheets.
Silk has virtually Thermally stable between 140°C to 175° C	

**Chemical Structure of Silk Fibroin :**



**Figure 1. Chemical Structure of Silk Fibroin.<sup>28</sup>**

**Procedure for Extraction of Fibroin and Sericin.**



**Figure 2.**Extraction of fibroin and sericin from the cocoons of *B.morisilkworms*.<sup>24,29</sup>

**Technique of Preparation for Silk Fibroin Based Nanoparticles.**<sup>30</sup>

Numerous methods are available for the formulation of SF-based nanoparticles or microspheres, like salting out, desolvation, electrospraying and

mechanical comminution, or PVA blend technique. The high protein nature and molecular weight of Silk Fibroin make the preparation of Nano formulation difficult to control. Furthermore Silk Fibroin tends to self-assemble into gels or fibers upon exposure to high shear, pH change, salt and heat.

**Table 3. Methods for Preparation of silk fibroin-based Nanoparticles.**

Sr. No.	Preparation Methods	Advantages	Disadvantages	Particle Size
1.	Salting out	Low cost; high yield; Avoids use of toxic solvents and safe operations	Salting out agents residue.	486~1200 nm
2.	PVA blend film method	Time and energy efficient. Mild operation conditions or safe to manipulate.	PVA residue.	500 nm~2 mm
3.	Desolvation	Small particle size;	Organic solvent residue	35~125 nm

		Simplicity of operation.		150~170 nm
4.	Electric fields	Mild operation conditions;	Particle with big size.	200 nm~3 $\mu$ m
5.	Electrospraying	Controllable particle size; Simplicity of operation.	To induce insolubility of SF	59~75 nm
6.	Mechanical Comminution	Easy to scale up	The impurities and any grinding aids need be removed	~200 nm

### Salting out

Salting out Method involves the salting out of protein solution to obtain protein co-acerbates. Proteins has hydrophobic and hydrophilic parts. Hydrophobic parts could be interrelated through the water molecules and permit the proteins to attained hydrogen bonds among the adjoining water molecules. By the increasing of the salt concentration the ions of salt attract a few of water molecules and as a resulting in the confiscation of the water fence among protein molecules. The protein molecules amassed together by formed hydrophobic exchanges with each other and the solution. Formation of Silk Fibroin nanoparticles with normal diameter of 486~1200 nm in aqueous development through salting out method.<sup>31,32</sup>

### Desolvation

Generally this technique is used to prepare protein-based nanoparticles. The desolvation or simple coacervation process decrease the solubility of the protein foremost to phase separation. The affixing of desolvating agent leads to conformation modify in protein structure ensuing in precipitation or coacervation of the protein. Throughout the phase separation and phase by means of a colloidal constituent or co-acervate and subsequent stage via solvent or non-solvent mixture are formed. A steady

particle size is reached after an original development stage that extra desolvation exclusively lead to enlarged element yield.<sup>33,34,35</sup>

### Mechanical Comminution

Comminution is the decrease of solid resources as of single usual particle size to a lesser average crushing, particle size, or grinding and milling *etc.* The method usually involves high energy wet or dry milling through the adding up of mill aids and naturally use milling period from several hours up to lots of days. The process is simple to function and scale up. Technique still experience from difficulty in ensure to all the particle be crushed properly. Extended mill time as well result in extra milling impurities. Furthermore, the constituent part size division is wide.<sup>36,37</sup>

### PVA Blend Film Method

Silk Fibroin constituent part with convenient particle size (500 nm~2 mm) as well as shape via PVA as a continuous phase to separate Silk Fibroin solution into microspheres or nanoparticles in Silk fibroin: PVA blend films at a weight ratio from 1/1 to 1/4. The procedure was based on phase separation between SF and polyvinyl alcohol (PVA). The SF/PVA blend solution have dried out into a film firstly. After that water insoluble Silk Fibroin particle

might be fabricated via film dissolution in water and subsequent centrifugation toward take away PVA. The process was easily because the process in used with PVA and water an FDA-approved substance. Through regulate the concentration of SF and PVA or take up ultra-sonication on the blend solution. The SF particles with diverse particle size can be prepared. Drug could be loaded into Silk Fibroin particle via addition model drugs in the original Silk Fibroin solution. These Silk Fibroin element contain probable as drug carriers in field of biomedical applications.<sup>38</sup>

**Electrospraying**

Electrospraying is a technique of liquid atomization via means of electrical *forces* with is an emerging process for the fast and elevated during put making of nanoparticles. In electros praying process, the liquid smooth out of a capillary nozzle, preserved at high electric potential and forced with the electric field to dispersed into small droplets.<sup>39</sup>

**Electric Fields**

In this process the arrangement of the Silk Fibroin nanoparticles through size of tens of nanometers is significant pace. Underneath electric pasture the

nanoparticles comprehensive to form microspheres or nanoparticle on top of positive electrodes put off inter molecular self-assembly of Silk Fibroin in impartial solution.<sup>40</sup>

**METHODS OF PREPARATION FOR SILK FIBROIN-BASED MICROSPHERES.**

The following method is used for preparation of silk micro-spheres which can be used to encapsulate and release growth factors with small molecules and therapeutic compounds.

**1. Prepared byDOPC Method<sup>41</sup>**

- ✓ Aqueous silk solution, 8% (wt/vol)
- ✓ 1,2-Dioleoyl-*sn*-glycero-3-phosphocholine (DOPC)
- ✓ Chloroform
- ✓ Methanol ,Nitrogen gas
- ✓ Drug, small molecule or protein of interest
- ✓ Ultrapure water

**2. Prepared by PVA Method<sup>42,43</sup>**

- ✓ Aqueous silk solution, 8% (wt/vol)
- ✓ PVA Polyvinyl Alcohol (MW 30–70 kDa;
- ✓ Ultrapure water

**APPLICATIONS OF SILKWORM<sup>44</sup>**

**Table No 4. Formulations and applications of Silkworm**

Sr.No	Application	Tissue type	Formulation
1.	Drug delivery	1. Drug Delivery 2. Small Molecules 3. Growth Factor delivery	1. Spheres 2. Mcro- or nanoparticle 3. Spheres
2.	Implant devices	1. Anterior cruciate ligament 2. Mandibular defects	1. Fibers 2. Aqueous sponges
3.	Disease models	1. Breast cancer	1. HFIP sponges



4	Tissue engineering	<ol style="list-style-type: none"> <li>1. Tissue engineering</li> <li>2. Corneal</li> <li>3. Cervical tissue</li> <li>4. Cartilage</li> <li>5. Skin</li> <li>6. Vascular tissue</li> </ol>	<ol style="list-style-type: none"> <li>1. HFIP sponges or Hydrogels</li> <li>2. Patterned silk films</li> <li>3. Aqueous sponges</li> <li>4. HFIP sponges or Hydrogels</li> <li>5. Electrospun fibers</li> <li>6. Tubes or Electrospun fibers</li> </ol>
---	--------------------	--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------	----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------

## APPLICATION

### Preparation of silkworm silk- based biomaterials

Silk has been used in biomedical sutures since its mechanical strength and biocompatibility. Though silk associated with pollutant sericin proteins is a probable allergen reason. The degradation invention of silk fibroin proteins among beta-sheet structures from the action of proteases like alpha-chymotrypsin have recently be details or no cytotoxicity was experiential to neuron cells *in-vitro*.<sup>45,46</sup>

### Films

Silk films have been used with different covalent decoration of functional peptides like implants for drug Delivery and bone formation. For parathyroid hormone (PTH), bone regeneration, RGD, and BMP-2, can be directly immobilized on silk films by carbodiimide chemistry. Differentiation of human bone marrow-derived stem cells cultured with the bedecked silk films can be induced by immobilized BMP-2. Moreover, the usefulness of silk films to support long-term adenosine release from adenosine kinase deficient embryonic stem cells has been examined. Silk films adorned with bioactive molecules can be used for local drug delivery through direct implantation.<sup>47,48</sup>

### Nanoparticles

Silk-based nanoparticles have been prepared in nature derived silk fibroin support nanoparticles less than 100 nm) for local and sustained therapeutic curcumin delivery toward cancer cells. Curcumin loaded silk nanoparticles prepared in various proportions with pure silk fibroin or silk fibroin with chitosan and PVA were spherical, stable, negatively charged, 150-170 nm in average diameter as well as showed no toxicity. The silk-based nanoparticles enclose curcumin demonstrate a higher effectiveness against breast cancer cells also contain potential to care for *in-vivo* breast tumors via local or sustained and long term therapeutic delivery.<sup>49,50</sup>

### Coatings

Coatings of silk fibroin contain studied to give interface for biomaterials. The forceful force of self-assembly to form coatings is electrostatic and some hydrophobic interactions. The elasticity of silk based coating have been investigated using an aqueous step wise evidence process with *B. mori*. The width of one layer was reported to be around 10 nm while put down from a one mg/mL aqueous solution of silk. The secondary arrangement of silk fibroin in the coating was regulated to manage the biodegradation rate, which indicates layer thickness and numbers of

layers and secondary structure of the layers.<sup>51</sup> Nano layer coatings of silk fibroin is used to enclose model compounds of small molecule drugs or therapeutically applicable proteins. Multilayered silk based coatings have been urbanized or used as drug carriers and delivery systems to estimate vascular cell comeback to heparin and clopidogrel.<sup>52,53</sup>

### Microspheres

Silk fibroin microspheres had been prepared via different techniques like spray-drying, desolvation, electric field though, the size of the microspheres had above 100 µm, which is suboptimal intended for drug delivery. Further techniques are prepared silk microspheres include lipid vesicles as templates to professionally load bioactive molecules used for local controlled releases be reported recently. The lipid is then removed by methanol or NaCl, resulting in silk microspheres consist of beta-sheet structure and around 2 µm in diameter.<sup>54,55</sup>

### Implants, tubes and scaffolds

Silk-based Three D scaffold loaded through one morphogenetic protein-2 (BMP-2) to encourage human bone marrow stromal cells to undertake estrogenic differentiation due to their biocompatibility and mechanical properties. Horseradish peroxidase (HRP) enzyme gradients

be immobilized on silk 3D scaffolds to arrange original functional scaffolds as well as regional patterning of the rise to manage cell and tissue outcomes currently studies show adenosine release through silk based implants to the brain have been studied for refractory epilepsy treatments.<sup>56,57</sup> Silk based implants releases adenosine demonstrated therapeutic ability or together with the controlled release of adenosine over a period of two weeks through slow degradation of silk or biocompatibility and the release of prearranged dose of adenosine. Nerve growth factor (NGF) loaded silk fibroin nerve encompass been studied to guide the developing of axons and physically defend the axonal cone for peripheral nerve repair. Silk fibroin scaffolds have insulin-like growth factor I (IGF-I) have been formulated for controlled IGF-I release in the situation of cartilage repair<sup>58</sup>

Tropical tasar silkworm *Antheraea mylitta* had also estimated for *in-vitro* drug release along with for the study of cell surface interactions. Silk based micro molded matrix hold up a significant development in cell attachment or spreading mitochondrial activity along with proliferation through feline fibroblasts in association to polystyrene plates as controls.<sup>59</sup>

**Table No 5. Silk Fibroin Patents**

Sr No.	Publication No	Inventors	Title	Reference
1.	EP2403551 A2	David L. Kaplan, Bruce Panilaitis, Eleanor M. Pritchard, Fiorenzo Omenetto, Jordan Axelrad	Silk fibroin systems for antibiotic delivery	60
2.	WO 2013159101 A1	Evangelia BELLAS, Amanda BARYSHYAN, Lindsay WRAY, David L.	Silk fibroin based care compositions	61

		Kaplan		
3.	US 20150164117	Kaplan, David L. (Concord, MA, US) Omenetto, Fiorenzo (Lexington, MA, US)	Encapsulation of fragrance and/or flavors in silk fibroin biomaterials	62
4.	US5252285 A	Robert L. Lock	Process for making silk fibroin fibers	63
5.	US8178656 B2	David L. Kaplan, Meinel Lorenz	Silkbased drug delivery system	64
6.	WO2013142119 A1	David L. Kaplan, Tuna Yucel, Michael L. Lovett, Xiaoqin Wang	Silk reservoirs for drug delivery	65

**Table No. 6 Silk fibroin based Marketed Formulation:**

Silk fibroin based formulation are marketed in mainly in China in Powder form.

Sl. no	Formulation	Brand Name	Type	Function	Business Type
1.	Powder	Silk Peptide Powder	Watersoluble polypeptide substance	Skin wound healing.	HuzhouXintiansi Biotech Co., Ltd. China
2.	Liquid	Silk Peptide Liquid	Water soluble polypeptide substance	Hair Care Function	China tiansi biotech co.,ltd, seoul korea
3.	Powder	Silk fibrin powder	macromolecular silk fibrin	natural moisturizing, absorb part of ultraviolet ray	HuzhouXintiansi Biotech Co., Ltd. China
4.	Powder	Silk fibroin powder	Solvent Extraction	nourishing skin	Xi'an Lyphar Biotech Co., Ltd, China.
5.	Silk fibroin powder	ZhongYun	Solvent Extraction	natural and promoting healing of skin wounds	Trading Company Shaanxi, China
6.	Amino acids silk protein	Lingeba	Silk Fibroin extract	Smoothness for natural hair	Hangzhou, zhejiang, China

**CONCLUSION**

Silk Fibroin is an attractive polymer for drug delivery of bioactive compounds for the sustained or controlled drug delivery. Silk Fibroin based matrixes, micro particles, nanoparticles were prepared from aqueous solutions were stable, negatively charged, spherical and showed no toxicity of the fibroin

protein polymer and prepared by the different techniques. Crystalline can be controlled and induced by action with different solvents. Drug release from these coating have sustained or controlled by number of layers, layer thickness and secondary arrangement of the silk fibroin layer.

**REFERENCES**

1. Debjit B, Harish G, Duraivel K, Sampath PK., Silk based drug delivery systems., The pharma innovation., 2013;11:42-48.
2. Shewry PR, Tatham AS, Bailey AJ., Elastomeric proteins: structures, biomechanical properties, and biological roles., Cambridge uni press., 2004: 136-174.
3. Yücel T, Kojic N, Leisk GG, Lo TJ, Kaplan DL., Non-equilibrium silk fibroin adhesives., J structure bio., 2010; 170: 406-412.
4. Kundu J, Chung YI, Kim YH, Tae G, Kundu Sc., Silk fibroin nanoparticles for cellular uptake and control release., Int J Pharm., 2010;388:242-250.
5. Vollrath F, Knight DP., Liquid crystalline spinning of spider silk., Nature., 2001;410: 541-548.
6. Numata k, Cebe P, Kaplan DL., Mechanism of enzymatic degradation of beta sheet crystals., Biomaterials., 2010;31:2926-2933.
7. Wang Y, Kim HJ, Vunjak NG, Kaplan DL., Stem cell-based tissue engineering with silk biomaterials., Biomaterials., 2006; 27: 6064-6082.
8. Makaya K, Terada S, Ohgo K, Asakura T., Comparative study of silk fibroin porous scaffolds derived from salt/water and sucrose/hexafluoroisopropanol in cartilage formation., J bioscibioeng., 2009;108:68-75.
9. Liu Y, Liu H, Qian J, Deng J, Yu T., Structure and properties of the composite membrane of regenerated silk fibroin and PVA and ITS application to amperometric tetrahydroamperometric glucose sensor., J MacromolSci Pure Appl Chem., 1996;33:209-219.
10. Guerette PA, Ginzinger DG, Weber BH, Gosline JM., Silk properties determined by gland-specific expression of a spider fibroin gene family., Sci., 1996;272:112-115.
11. Winkler S, Wilson D, Kaplan DL., Controlling beta-sheet assembly in genetically engineered silk by enzymatic phosphorylation/dephosphorylation., Biochem., 2000;39:12739-12746.
12. BiramSaheb NM, Singh T, Beera S., Occurrence of Unfertilized Eggs in the Mulberry Silkworm, *Bombyx mori* (L.) (Lepidoptera: Bombycidae), Int J IndustEntomol., 2009 ;18(1):1-7.
13. Jiang CY, Wang XY, Gunawidjaja R, Lin Yh, Gupta Mk, Kaplan DL, et al., Mechanical properties of robust ultrathin silk fibroin films., Advfunct mater., 2007;17:2229-2237.

14. Yokoyama T, Smith RE, Mittler TE, Smith CN. Annual Reviews.,Palo Alto California .,1973;267-284
15. AnubhavN,RajeevKS.,Applications of silk in drug delivery: advancement in pharmaceutical dosage forms., Indo Global Journal of Pharma Sci., 2013; 3(3): 204-211.
16. Szybala C, Pritchard EM, Lusardi TA, LI T, Wilz A, Kaplan DL, Boison D., Antiepileptic effects of silk-polymer based adenosine release in kindled rats., Exp Neurol., 2009;219:126–135.
17. Lazaris A, Arcidiacono S., Spider silk fibers spun from soluble recombinant silk produced in mammalian cells., Sci.,2002;295:472–476.
18. Hinman MB, Lewis RV.,Isolation of a clone encoding a second dragline silk fibroin., J biol chem., 1992;267:19320–19324.
19. Akai H, Nagashima T., Fine-structural characteristics of anaphe cocoon filament., Int J. Wild silk moth silk., 1999;4:13–16.
20. Eisoldt L, Hardy J, Heim M, Scheibel T., The role of salt and shear on the storage and assembly of spider silk proteins., J struct biol., 2010;170:420-425.
21. Numata K, Hamasaki J, Subramanian B, Kaplan DL., Gene delivery mediated by recombinant silk proteins containing cationic and cell binding motifs., In preparation J control release., 2010; 146(1): 136-43.
22. Hu X, Kaplan DL., silk biomaterials. In: editor-in-chief: paul d, editor.comprehensivebiomaterials., Oxford elsevier., 2011;212;207-19.
23. Okazaki Y, Tomotake H, Tsujimoto K, Sasaki M, Kato N., Consumption of a resistant protein, sericin, elevates fecal immunoglobulin a, mucins, and cecal organic acids in rats fed a high-fat diet., J Nutrition., 2011;141(11):1975-81
24. Pritchard EM, Kaplan DL., Silk fibroin biomaterials for controlled release drug delivery., Expert opinion on drug delivery .,2011;8(6):797-811
25. Hazeri N, Tavanai H, Moradi AR., Production and properties of electro sprayed sericinnano powder.,Sci& tech adv materials., 2012;13(3):035010
26. Huang D, Wang L, Dong Y, Pan X, Li G, Wu C., A novel technology using transcleral ultrasound to deliver protein loaded nanoparticles., Eur J Pharm Biopharma., 2014;88: 104–115.
27. Foo CW, Bini E, Hensman J, Knight D, Lewis RV, Kaplan DL., Role of ph and charge on silk protein assembly in insects and spiders., appl. Phys., 2006;82:223–233.
28. [http://wwwchem.uwimona.edu.jm/courses/chem2402/textiles/silk\\_fibroin.gif](http://wwwchem.uwimona.edu.jm/courses/chem2402/textiles/silk_fibroin.gif)
29. Sobajo C, Behzad F, Yuan XF., Silk a potential medium for tissue engineering., Eplasty.,2008;8(47):438-446.
30. Zheng Z, YI L, Mao-bin X., Silk fibroin-based nanoparticles for drug delivery.,Int j mol sci., 2015;16:4880-4903.
31. Lammel As, Hu X, Park SH, Kaplan DL, Scheibel TR., Controlling silk fibroin particlefeatures for drug delivery.,Biomaterials., 2010;31:4583–4591.
32. Kumari A, YadavSk, Yadav SC., Biodegradable polymeric nanoparticles

- based drug delivery systems., *Colloids surf.*, 2010;75:1–18.
33. Kundu J, Chung YI, Kim YH, Tae G, Kundu SC., Silk fibroin nanoparticles for cellular uptake and control release., *Int. J. Pharm.*, 2010;388:242–250.
  34. Lohcharoenkal W Wang L, Chen YC, Rojanasakul Y., Protein nanoparticles as drug delivery carriers for cancer therapy., *Biomed. Res. Int.*, 2014;2014:180549.
  35. Pinto RC, Neufeld RJ, Ribeiro AJ, Veiga F., Nanoencapsulation : Methods for preparation of drug-loaded polymeric nanoparticles., *Nanomedicine.*, 2006;2:8–21.
  36. Tsuzuki T., Commercial scale production of inorganic nanoparticles., *Int. J. Nanotechnol.*, 2009;6:567–578.
  37. Koch CC., Top-down synthesis of nanostructured materials: mechanical and thermal processing methods., *Rev. Adv. Mater. Sci.*, 2003;5:91–99.
  38. Dai L, Li J, Yamada E., Effect of glycerin on structure transition of PVA/SF blends., *J Appl Polym Sci.*, 2002;86:2342–2347.
  39. Gholami A, Tavanai H, Moradi AR., Production of fibroin nanopowder through electrospaying., *J Nanopart Res.*, 2011;13:2089–2098.
  40. Lu Q, Huang Y, Li M, Zuo B, Lu S, Wang J, et al., Silk fibroin electro gelation mechanisms., *Acta Biomater.*, 2011;7:2394–2400.
  41. Wang X, Wenk E, Matsumoto A, Meinel L, Li C, Kaplan DL., Silk microspheres for encapsulation and controlled release., *J Control release.*, 2007;117:360–370.
  42. Wang X, Yucel T, Lu Q, Hu X, Kaplan DL., Silk nanospheres and microspheres from silk/pva blend films for drug delivery., *Biomaterials.*, 2010;31:1025–1035
  43. Altman GH, Diaz F, Jakuba C, Calabro T, Horan RL, Chen J, et al., Silk-based biomaterials., *Biomaterials.*, 2003;24:401–416.
  44. Murphy AR, Kaplan DL., Biomedical applications of chemically modified silk fibroin., *J Mat Chem.*, 2009; 19: 6443–6450.
  45. Hu X, Kaplan DL, Cebe P., Determining beta-sheet crystallinity in fibrous proteins by thermal analysis and infrared spectroscopy., *Macromolecules.*, 2006;39:6161–6170.
  46. Gobin AS, Rhea R, Newman RA, Mathur AB., Silk-fibroin-coated liposomes for long-term and targeted drug delivery., *Int j nanomed.*, 2006;1:81–87.
  47. Jin HJ, Park J, Valluzzi R, Cebe P, Kaplan DL., Biomaterial films of Bombyxmori silk fibroin with poly(ethylene oxide)., *Biomacromolecules.*, 2004;5:711–717.
  48. Uebersax L, Fedele DE, Schumacher C, Kaplan DL, Merkle HP, Boison D, . The support of adenosine release from adenosine kinase deficient cells by silk substrates., *Biomaterials.*, 2006;27:4599–4607.
  49. Mandal BB, Kundu SC., Self assembled silk sericin/poloxamer nanoparticles as nano carriers of hydrophobic and hydrophilic drugs for targeted delivery., *Nanotechnology.*, 2009;20:355101–355111.
  50. Lee KE, Cho SH, Lee HB, Jeong SY, Yuk SH., Microencapsulation of lipid nanoparticles containing lipophilic drug., *J microencapsul.*, 2003;20:489–496.

51. Yamaura K, Kuranuki N, Suzuki M, Tanigami T, Matsuzawa S., Properties of mixtures of silk fibroin/syndiotacti-rich poly (vinyl alcohol)., *J applpolym sci.*, 1990;41:2409–25
52. Wang X, Wenk E, Hu X, Castro GR, Meinel I, Li C, Merkle H, Kaplan DL., Silk coatings on plga and alginate microspheres for protein delivery., *Biomaterials.*, 2007;28:4161–4169.
53. Vepari CP, Kaplan DL., Covalently immobilized enzyme gradients within three-dimensional porous scaffolds.,*Biotechnolbioeng* .,2006;93:1130–1137.
54. Hino T, Shimabayashi S., Silk microspheres prepared by spray-drying of an aqueous system., *Pharmacy pharmacolcommun.*, 2000;6:335–39.
55. Goraltchouk A, Scanga V, Morshead CM, Shoichet MS., Incorporation of protein-eluting microspheres into biodegradable nerve guidance channels for controlled release., *J Control Release.*, 2006;110:400–407.
56. Uebersax L, Merkle HP, Meinel L., Insulin-like growth factor i releasing silk fibroin scaffolds induce chondrogenic differentiation of human mesenchymal stem cells., *J Control Release.*, 2008;127:12–21.
57. Zhang J, Pritchard E, Hu X, Valentin T, Panilaitis B, Omenetto FG, et al., Stabilization of vaccines and antibiotics in silk and eliminating the cold chain., *Proceedings of the national academy of sciences.* ,2012;109(30):11981-6.
58. Moisenovich MM, Pustovalova O, Shackelford J, Vasiljeva TV, Druzhinina TV, Kamenchuk YA, et al., Tissue regeneration in vivo within recombinant spidroin scaffolds., *Biomaterials.*, 2012;33(15):3887-98.
59. Kim UJ, Park J, Joo Kim H, Wada M, Kaplan DL., Three-dimensional aqueous-derived biomaterial scaffolds from silk fibroin., *Biomaterials.*, 2005;26(15):2775-2785.
60. David L., Silk fibroin systems for antibiotic delivery., *Ep2403551*;2012.
61. Evangelia B., Silk fibroin based personal care compositions.,*Wo 2013159101*;2013.
62. Kaplan D., Encapsulation of fragrance and/or flavors in silk fibroin biomaterials., *Us 20150164117* ;2015.
63. Robert L., Process for making silk fibroin fibers., *Us5252285*;1993.
64. David L., Silk based drug delivery system., *Us8178656 b2* ;2012.
65. David L., Silk reservoirs for drug delivery., *wo2013142119*;2013.

Conflict of Interest Reported: Nil;

Source of Funding: None Reported